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International Journal of Computational and Experimental Science and ENgineering (IJCESEN)

Vol. 11-No.3 (2025) pp. 5080-5093 <u>http://www.ijcesen.com</u>



Research Article

The Effect of ethanol extract of Kaffir lime leaves on Lipid Profile, Aterogenetic Index and Blood Vessel Histopathology of Male White Rats Hyperlipidemia Model

Reni Apriza^{1*}, Jekson Martiar Siahaan², Endy Juli Anto³, Syafruddin Ilyas⁴, Menang Bastanta Tarigan⁵

 ¹Master's Study Program in Biomedical Sciences, Faculty of Medicine, Universitas Methodist, Medan-Indonesia * Corresponding Author Email: reniapriza11@gmail.com - ORCID: 0000-0002-5247-7849
 ²Master's Study Program in Biomedical Sciences, Faculty of Medicine, Universitas Methodist, Medan-Indonesia, Email:siahaa2n@gmail.com- ORCID: 0000-0002-5247-7851
 ³Master's Study Program in Biomedical Sciences, Faculty of Medicine, Universitas Methodist, Medan-Indonesia, Email: ant2o@gmail.com - ORCID: 0000-0002-5247-7852
 ⁴Master's Study Program in Biomedical Sciences, Faculty of Medicine, Universitas Methodist, Medan-Indonesia, Email: ilya2s@gmail.com - ORCID: 0000-0002-5247-7853
 ⁵Master's Study Program in Biomedical Sciences, Faculty of Medicine, Universitas Methodist, Medan-Indonesia, Email: ilya2s@gmail.com - ORCID: 0000-0002-5247-7853
 ⁵Master's Study Program in Biomedical Sciences, Faculty of Medicine, Universitas Methodist, Medan-Indonesia, Email: ilya2s@gmail.com - ORCID: 0000-0002-5247-7853

Article Info:

Abstract:

DOI: 10.22399/ijcesen.3339 **Received:** 08 May 2025 **Accepted:** 05 July 2025

<u>Keywords</u>

Herbal Medicine, Lipids, Atherogenic Index, Histopathology, Kaffir Lime Leaves.

Kaffir lime leaf extract has the potential to be used as an antioxidant and antiinflammatory, apart from that, the compounds contained in kaffir lime leaves have properties that can be used as medicine. This study used a laboratory experimental design on male white Wistar hyperlipidemic rats. The research method uses a post-test only controlled group design. Hypercholesterolemia induction uses a high-fat feed method, this feed contains quail egg yolk, duck egg yolk, 3 mL/300 mg/kgBW/day of used cooking oil. The statistical tests used are data normality and homogeneity tests. The results of phytochemical screening on kaffir lime leaves showed secondary metabolites such as flavonoids, phenols, saponins, tannins and alkaloids. In this study there was a significant difference in mean body weight (p < 0.05) on the 14th day but there was no difference between groups on the 21st day. Apart from that, there were no significant differences in mean levels of total cholesterol, triglycerides, HDL (High-density lipoprotein), and LDL (Low-Density Lipoprotein) (p<0.05) between all groups. In eNOS levels there were no significant differences (p<0.05) between all groups. However, in HMGCOA levels, there were significant differences in levels (p<0.05) between all groups. Apart from that, the group with the ethanol extract of kaffir lime leaves at a dose of 35 mg/kgBW was better at controlling body weight, reducing total cholesterol, HDL, LDL and triglyceride levels, increasing eNOS levels and reducing HMGCOA levels.

1. Introduction

Hyperlipidemia is a condition where blood fat levels increase [1]. This is characterized by an increase in cholesterol, LDL (Low Density Lipoprotein) and triglyceride levels in the blood that exceed normal limits and a decrease in HDL (High Density Lipoprotein) [2, 3]. High cholesterol levels are a condition when the total blood cholesterol value increases above the normal value (>240mg/dL) (Dash & Saini, 2023). Where the limit for normal cholesterol levels in humans is <200mg/dL [4].

Overall, hyperlipidemia causes death in 4.4 million people in the world and causes 40.4 million

Disability Adjusted Life Years (DALY) [5]. Hyperlipidemia itself can occur if disorders of lipid and cholesterol metabolism are found, consisting of total LDL (Low-Density Lipoprotein) and HDL (High-density lipoprotein) cholesterol [6]. This increase in total cholesterol levels can increase risk factors for Coronary Heart Disease (CHD) and other cardiovascular diseases [7]. High total cholesterol levels will form atherosclerosis [8]. Atherosclerosis can cause hypertension and blockages in the blood vessels of the brain, heart and leg blood vessels [9]. One of the pathogeneses that causes atherosclerosis in blood vessel walls is an increase in fat levels in blood plasma [10,11]. So, it can cause blockages in the blood vessels of the heart or known as coronary heart disease (CHD) and stroke [12]. These two diseases are the biggest causes of death in the world [13].

Data from the World Health Organization (WHO) in 2014 shows that hyperlipidemia causes 18% of cardiovascular disease and 56% of ischemic heart disease worldwide [14, 5, 15]. Research results from the Health Research and Development Agency of the Ministry of the Republic of Indonesia in 2018 showed data that 1.5% or more than one million Indonesians had coronary heart disease [16]. Apart from that, 36.55% of Indonesians aged 15 years and over have cholesterol levels above normal (> 240 mg/dL), 24.9% have high LDL (Low Density Lipoprotein) levels (> 190 mg/dL), with HDL levels (High -density lipoprotein) below 40 mg/dl 24.3% and very high triglycerin levels (> 500 mg/dL) [17, 18].

In general, prevention of hyperlipidemia can be done by changing lifestyle for the better, such as maintaining diet and exercising [19, 20]. Meanwhile, pharmacological treatment of hyperlipidemia in most cases is conventional treatment [21]. Theoretically, preventive measures can be taken by administering medication [22]. However, continuous use of medication is not recommended [23]. Because several interventions originating from pharmacology are still doubtful about their results in reducing total cholesterol levels [24].

Apart from using conventional medicine, people also use traditional medicine as therapy for hyperlipidemia [25]. Research results in several health services in 7 provinces in Indonesia showed that 62.9% were given single herbal drug therapy and the rest were given a combination of herbal medicines and synthetic drugs [26]. It is hoped that the use of herbal medicines can reduce the use of synthetic drugs. Although the use of herbal medicines cannot completely replace the effects of synthetic drugs, herbal medicines are quite economical and have fewer side effects compared to synthetic drugs for treating hyperlipidemia in general [27].

Many medicinal plants have been proven to inhibit the formation and lower cholesterol, such as kaffir lime leaves [28]. Kaffir lime leaves contain flavonoid and phenolic compounds which act as antioxidants [29]. The flavonoid compounds contained work selectively on cholesterol metabolism in the liver in reducing cholesterol levels [30]. Flavonoids are able to increase bile acid excretion and reduce blood viscosity which causes cholesterol deposition in blood vessels [31]. This active ingredient has the potential to reduce total cholesterol levels in the blood [32].

Flavonoids, especially quercetin, tannins and saponins, are compounds that can reduce cholesterol levels [33]. Quercetin reduces cholesterol levels by reducing the α -tocopherol content which is part of LDL, the reduction that occurs results in inhibited LDL oxidation [34. 35]. The results of research by Rustanti & Lathifah [36], stated that giving quercetin extract can reduce total cholesterol levels. The greater the concentration of quercetin, the higher the anti-cholesterol activity.

The quercetin contained in kaffir lime leaves is a source of antioxidants that can ward off free radicals [37]. Saponin can reduce cholesterol levels and increase the binding of cholesterol to fiber, causing cholesterol in the intestine to be blocked and form complex bonds that are not soluble in fat [38, 39]. Tannins reduce cholesterol levels by reacting with mucosal proteins and epithelial cells which causes fat absorption in the intestine to be hampered [40]. Based on the explanations, opinions and previous research that have been stated in the paragraph above, the researchers are interested in researching more deeply regarding the influence of the ethanol extract of Kaffir lime on the lipid profile, atherogenic index and blood vessel histopathology of male white mice in the hyperlipidemia model.

2. Material and Methods

This research used a laboratory experimental design on male white Wistar hyperlipidemic rats. The research method uses a post test only controlled group design. This design allows researchers to measure the effect of treatment (intervention) on the experimental group by comparing the experimental group with the control group. In this design the researcher does not determine how big the change occurs, because the test is carried out at the end of the treatment.

Hypercholesterolemia induction uses a high-fat feed method, this feed contains quail egg yolk, duck egg volk, 3 mL/300 mg/kgBW/day of used cooking oil. The test animals were divided into 6 groups, namely the normal group (K1) which was not given treatment, (K2) which was given high fat feed 3 mL/200 mg/kgBW/day. (K3) were given high fat feed 3 mL/200 mg/kgBW/day + simvastatin 2.1 mg/kgBW/day. (K4) were given high-fat feed 3 mL/300 mg/kgBW/day + ethanol extract of kaffir lime leaves at a dose of 35 mg/200gBW. (K5) was given a high fat diet of 3 mL/300 mg/kgBW/day + ethanol extract of kaffir lime leaves at a dose of 70 mg/200gBW. (K6) was given high-fat feed 3 mL/300 mg/kgBW/day + ethanol extract of kaffir lime leaves at a dose of 140 mg/200gBW [41, 15].

The research was carried out at the Biomedical Science Master's Laboratory, Faculty of Medicine,

Indonesian Methodist University to produce ethanol extract from kaffir lime and the animal house as a place for surgical treatment of experimental animals. Faculty of Medicine, Master of Biomedical Sciences, Indonesian Methodist University, for examination of kaffir lime characterization and screening, examination related to lipid profile (total cholesterol, HDL cholesterol, LDL cholesterol and ratio of total cholesterol/HDL cholesterol), atherogenic index, and blood vessel histopathology. The statistical tests used are data normality and homogeneity tests. The data was found to be normally distributed so an ANOVA test was carried out. All data analysis was carried out using SPSS software. In this study, for statistical test decisions, a real level of 5% (p = 0.05) was taken which was considered meaningful or significant. If a p value <0.05 is obtained then a follow-up post hoc test is carried out.

3. Results and Discussions

Results

A. Total Flavonoid Test

The total flavonoid content was measured using a sample test with a concentration of 10 ppm, 0.5 mL was put into a 5 mL measuring flask, then 0.1 mL of 2% AlCl3, 0.1 mL of 1 M CH3COONa were added, then incubated for 30 minutes, absorbance of the sample solution was measured using a UV-Vis spectrophotometer with a maximum wavelength of 415 nm. The total flavonoid content was calculated using a linear regression equation from the obtained quercetin standard curve. The results of measuring the standard absorbance of quercetin can be seen in Figure 1 and the total flavonoids of the ethanol extract of kaffir lime leaves can be seen in Table 1.

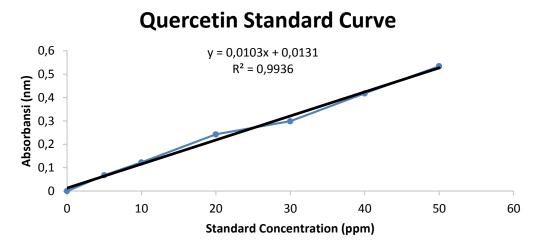


Figure 1. Quercetin standard curve

Sample	Extract Concentration (ppm)	Replication	Absorbance (nm)	Total Flavonoid Level (mg/L)	Average Total Flavonoid Levels (mg/L)
Kaffir Lime Leaf Ethanol Extract	10	1	0.2773	25.650	
		2	0.2874	26.631	26.417
		3	0.2909	26.971	

Table 1. Total Flavonoids from Ethanol Extract of Kaffir Lime Leaves

Table 2. Total Phenolics Ethanol Extract of Kaffir Lime Le	eaves
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Sample	Extract Concentration (ppm)	Replication	Absorbance (nm)	Total Phenolic Content (mg/L)	Average Total Phenolic Content (mg/L)
Kaffir Lime		1	0.342	484.957	
Leaf Ethanol	110	2	0.356	504.857	503.429
Extract		3	0.367	520.571	

B. Total Phenolic Test

The total phenolic content was measured using a sample test with a concentration of 110 ppm, 0.5 mL was put into a 5 mL measuring flask, then 0.1 mL of Follin-Ciocalteau reagent was added, shaken and left

for 4-8 minutes. Then 0.1 mL of 2% Na2CO3 was added and filled with aquaset to the limit mark. After incubation for 30 minutes, the absorbance of the sample solution was measured with a UV-Vis spectrophotometer at a maximum wavelength of 765 nm. The total phenolic content was calculated using a linear regression equation from the obtained gallic acid standard curve. The results of measuring the standard absorbance of gallic acid can be seen in Figure 2 and the total phenolics of the ethanol extract of kaffir lime leaves can be seen in Table 2.

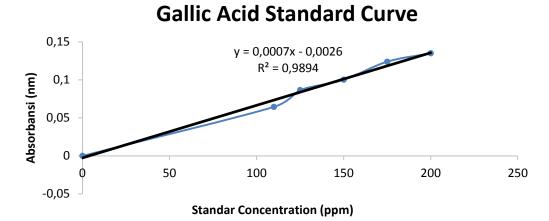


Figure 2. Gallic acid standard curve

C. Phytochemical Screening Analysis of Ethanol Extract of Kaffir Lime Leaves

The results of the phytochemical screening examination of the ethanol extract of kaffir lime

leaves showed secondary metabolites such as flavonoids, alkaloids, saponins, tannins and phenols. The results of the screening examination can be seen in Table 3.

Table 3. Phytochemical Screening of Ethanol Extract of Kaffir Lime Leaves

Test	Flavonoids	Alkaloids	Saponins	Tannin	Phenol
Picture			and a		B St van
Results	(+)	(+)	(+)	(+)	(+)

D. Mean Value of Differences in Body Weight After Giving Ethanol Extract of Kaffir Lime Leaves

Wistar white rats (Rattus norvegicus) with a hyperlipidemia model after administration of kaffir lime leaf ethanol extract can be seen in Table 4.

The results of the analysis of differences in body weight levels H-0, H-7 and H-14 in a group of male

Table 4. Results of Analysis of Differences in Body Weight Levels on H-0, D-7 and D-14 in Groups After Giving Ethanol

 Extract of Kaffir Lime Leaves

PostHoc		BB H-0	BB H-7	BB H-14
		Р	Р	Р
	Negative Group	< 0.001*	< 0.001*	0.002*
	Positive Group	< 0.001*	< 0.001*	0.330*
Normal Group	Group I	< 0.001*	0.005*	0.534*
1	Group II	< 0.001*	< 0.001*	0.289*
	Group III	< 0.001*	0.027*	0.964*
	Normal Group	< 0.001*	< 0.001*	0.002*
	Positive Group	0.751	0.076*	0.026*
Negative Group	Group I	0.313	0.002*	0.011*
	Group II	0.657	0.030*	0.032*
	Group III	0.150	< 0.001*	0.003*
	Normal Group	< 0.001*	< 0.001*	0.330*
Desitive Crown	Negative Group	0.751	0.076*	0.026*
Positive Group	Group I	0.468	0.121*	0.721*
	Group II	0.447	0.667*	0.929

PostHoc		BB H-0	BB H-7	BB H-14
PO	sthoc	Р	Р	Р
	Group III	0.082	0.030*	0.352
	Normal Group	< 0.001*	0.005*	0.534*
	Negative Group	0.313	0.002*	0.011*
Group I	Positive Group	0.468	0.121	0.721*
1	Group II	0.150	0.255	0.656*
	Group III	0.018	0.504	0.563*
	Normal Group	< 0.001*	< 0.001*	0.289*
	Negative Group	0.657	0.030*	0.032*
Group II	Positive Group	0.447	0.667*	0.929*
	Group I	0.150	0.255*	0.656*
	Group III	0.313	0.076*	0.309*
	Normal Group	<0.001*	0.027*	0.964*
	Negative Group	0.150	< 0.001*	0.003*
Group III	Positive Group	0.082	0.030*	0.352*
-	Group I	0.018	0.504*	0.563*
	Group II	0.313	0.076*	0.309*

*Post Hoc (Significant <0.05)

E. Mean Value of Differences in Total Cholesterol Levels After Giving Ethanol Extract of Kaffir Lime Leaves

The results of the analysis of differences in total cholesterol levels on H-0, H-7, H-14 and H-21 in a

group of male white Wistar rats (Rattus norvegicus) with a hyperlipidemia model after administration of kaffir lime leaf ethanol extract can be seen in Table 5.

Table 5. Results of Analysis of Differences in Total Cholesterol Levels on H-0, H-7, H-14 and H-21 in Groups After Giving Ethanol Extract of Kaffir Lime Leaves

Total Cholesterol Levels		H-0	D-7	H-14	H-17
		Р	Р	Р	Р
Normal Group	Negative Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*
	Positive Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*
	Group I	< 0.001*	< 0.001*	< 0.001*	< 0.001*
	Group II	< 0.001*	< 0.001*	< 0.001*	< 0.001*
	Group III	< 0.001*	< 0.001*	< 0.001*	< 0.001*
Negative Group	Normal Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*
	Positive Group	0.002	0.015*	< 0.001*	0.228*
	Group I	0.062	0.238*	0.013*	0.001*
	Group II	0.314	0.575*	0.532*	< 0.001*
	Group III	0.500	0.916*	0.814*	< 0.001*
Positive Group	Normal Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*
	Negative Group	0.002*	0.015*	< 0.001*	0.228*
	Group I	0.182*	0.177*	0.024*	0.018*
	Group II	0.030*	0.052*	< 0.001*	0.005*
	Group III	0.014*	0.011*	< 0.001*	0.002*
Group I	Normal Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*
	Negative Group	0.062*	0.238*	0.013*	0.001*
	Positive Group	0.182*	0.177*	0.024*	0.018*
	Group II	0.370*	0.528*	0.053*	0.606*
	Group III	0.220*	0.200*	0.022*	0.376*
Group II	Normal Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*
	Negative Group	0.314*	0.575*	0.532*	< 0.001*
	Positive Group	0.030*	0.052*	< 0.001*	0.005*
	Group I	0.370*	0.528*	0.053*	0.606*
	Group III	0.735*	0.506*	0.695*	0.708*
Group III	Normal Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*
	Negative Group	0.500*	0.916*	0.093	< 0.001*
	Positive Group	0.014*	0.011*	0.415	0.002*
	Group I	0.220*	0.200*	0.038	0.376*
	Group II	0.735*	0.506*	0.181	0.708*

*Post Hoc (Significant <0.05)

F. Mean Values of Differences in Plasma Atherogenic Index (IAP) Levels After Administration of Kaffir Lime Leaf Ethanol Extract

The results of the analysis of differences in plasma atherogenic index (IAP) levels between groups of male Wistar white rats (Rattus norvegicus) with a hyperlipidemia model after administration of kaffir lime leaf ethanol extract can be seen in Table 6.

 Table 6. Results of Analysis of Differences in Plasma Atherogenic Index (IAP) Levels Between Groups After

 Administration of Kaffir Lime Leaf Ethanol Extract

Test	Crown	IAP	
	Group	Mean ± SD	Р
Pre	Normal Group	0.430±0.259	
	Negative Group	0.220±0.038	
	Positive Group	0.236±0.025	<0.001*
	Group I (100)	0.220±0.018	
	Group II (200)	0.220±0.015	
	Group III (300)	0.220±0.015	
Post	Normal Group	0.065±0.037	< 0.001*
	Negative Group	0.241±0.021	
	Positive Group	0.151±0.027	
	Group I (100)	0.219±0.020	
	Group II (200)	0.199±0.034	
	Group III (300)	0.187±0.028	

*Anova Test (Significant < 0.05)

G. Mean Value of Differences in Total Cholesterol Levels Pre and Post Triglyceride Levels After Administration of Kaffir Lime Leaf Ethanol Extract

The results of the analysis of differences in total cholesterol levels and pre and post triglyceride levels

in a group of male Wistar white rats (Rattus norvegicus) with a hyperlipidemia model after administration of kaffir lime leaf ethanol extract can be seen in Table 7.

Table 7. Results of Analysis of Differences in Total Cholesterol Levels and Triglyceride Levels Pre and Post in Groups

 After Giving Ethanol Extract of Kaffir Lime Leaves

		Total ch	olesterol	Triglyceride Levels		
Group Normal Group Negative Group		Pre	Post	Pre	Post	
		Р	Р	Р	Р	
Normal Group	Negative Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*	
-	Positive Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*	
	Group I	< 0.001*	< 0.001*	< 0.001*	< 0.001*	
	Group II	< 0.001*	< 0.001*	< 0.001*	< 0.001*	
	Group III	< 0.001*	< 0.001*	< 0.001*	< 0.001*	
Negative Group	Normal Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*	
	Positive Group	0.004*	< 0.001*	0.033*	0.015*	
	Group I	0.080*	0.004*	0.791*	0.107*	
	Group II	0.345*	0.014*	0.285*	0.333*	
	Group III	0.500	0.033*	0.413*	0.563*	
Positive Group	Normal Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*	
_	Negative Group	0.004*	< 0.001*	0.033*	0.015*	
	Group I	0.210*	0.015*	0.072*	0.364*	
	Group II	0.041*	0.004*	0.002*	0.120*	
	Group III	0.020*	0.001*	0.005*	0.055*	
Group I	Normal Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*	
	Negative Group	0.080*	0.004*	0.791*	0.107*	
	Positive Group	0.210*	0.015*	0.072*	0.364*	
	Group II	0.400*	0.597*	0.157*	0.504*	
	Group III	0.250*	0.363*	0.242*	0.290*	
Group II	Normal Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*	

			Total cholesterol		ride Levels
Group		Pre	Post	Pre	Post
		Р	Р	Р	Р
	Negative Group	0.345*	0.014*	0.285*	0.333*
	Positive Group	0.041*	0.004*	0.002*	0.120*
	Group I	0.400*	0.597*	0.157*	0.504*
	Group III	0.751*	0.700*	0.797*	0.692*
Group III	Normal Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*
	Negative Group	0.527*	0.033*	0.413*	0.563*
	Positive Group	0.020*	0.001*	0.005*	0.055*
	Group I	0.250*	0.363*	0.242*	0.290*
	Group II	0.751*	0.700*	0.797*	0.692*

*Anova Test (Significant < 0.05)

H. Mean Value of Differences in HDL Levels and LDL Levels Pre and Post After Giving Ethanol Extract of Kaffir Lime Leaves The results of the analysis of differences in HDL

levels and pre and post LDL levels in a group of male

Wistar white rats (Rattus norvegicus) with a hyperlipidemia model after administration of kaffir lime leaf ethanol extract can be seen in Table 8.

Table 8. Results of Analysis of Differences in HDL and LDL Levels Pre and Post in Groups After Giving Ethanol

 Extract of Kaffir Lime Leaves

		HD	DL levels	LD	L levels
Group		Pre	Post	Pre	Post
		Р	Р	Р	Р
Normal Group	Negative Group	< 0.001*	0.026*	< 0.001*	< 0.001*
	Positive Group	< 0.001*	< 0.001*	< 0.001*	0.048*
	Group I	< 0.001*	0.011*	< 0.001*	< 0.001*
	Group II	< 0.001*	< 0.001*	< 0.001*	< 0.001*
	Group III	< 0.001*	< 0.001*	< 0.001*	< 0.001*
Negative Group	Normal Group	< 0.001*	0.026*	< 0.001*	< 0.001*
	Positive Group	0.568*	< 0.001*	0.064*	< 0.001*
	Group I	0.927*	0.731*	0.320*	0.011*
	Group II	0.525*	0.017*	0.142*	0.001*
	Group III	0.649*	0.001*	0.317*	< 0.001*
Positive Group	Normal Group	< 0.001*	< 0.001*	< 0.001*	0.048*
-	Negative Group	0.586*	< 0.001*	0.064*	< 0.001*
	Group I	0.525*	< 0.001*	0.367*	< 0.001*
	Group II	0.927*	0.011*	0.679*	< 0.001*
	Group III	0.927*	0.175*	0.371*	0.002*
Group I	Normal Group	< 0.001*	0.011*	< 0.001*	< 0.001*
-	Negative Group	0.927*	0.731*	0.142*	0.011*
	Positive Group	0.525*	< 0.001*	0.679*	< 0.001*
	Group II	0.468*	0.038*	0.622*	0.399*
	Group III	0.586*	0.002*	0.627*	0.203*
Group II	Normal Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*
-	Negative Group	0.525*	0.017*	0.142*	0.001*
	Positive Group	0.927*	0.011*	0.679*	< 0.001*
	Group I	0.468*	0.038*	0.622*	0.399*
	Group III	0.856*	0.202*	0.627*	0.659*
Group III	Normal Group	< 0.001*	< 0.001*	< 0.001*	< 0.001*
-	Negative Group	0.649*	0.001*	0.317*	< 0.001*
	Positive Group	0.927*	0.175*	0.371*	0.002*
	Group I	0.586*	0.002*	0.994*	0.203*
	Group II	0.856*	0.202*	0.627*	0.659*

*Anova Test (Significant < 0.05)

İ. Mean Value of Differences in HMGCOA Levels Pre and Post After Giving Kaffir Lime Leaf Ethanol Extract

white rats (Rattus norvegicus) with a hyperlipidemia model after administration of kaffir lime leaf ethanol extract can be seen in Table 9.

The results of the analysis of differences in pre and post HMGCOA levels in a group of male Wistar

Table 9. Results of Analysis of Differences in Pre and Post HMGCOA Levels in Groups After Administration of Kaffir	
Lime Leaf Ethanol Extract	

HMGCOA levels		Pre	Post
		Р	Р
Normal Group	Negative Group	<0.001*	<0.001*
	Positive Group	<0.001*	0.048*
	Group I	<0.001*	<0.001*
	Group II	<0.001*	<0.001*
	Group III	<0.001*	<0.001*
Negative Group	Normal Group	<0.001*	<0.001*
	Positive Group	<0.001*	<0.001*
	Group I	0.022*	0.011*
	Group II	0.029*	0.001*
	Group III	0.029*	<0.001*
Positive Group	Normal Group	<0.001*	0.048*
	Negative Group	<0.001*	<0.001*
	Group I	0.004*	<0.001*
	Group II	0.003*	<0.001*
	Group III	0.003*	0.002*
Group I	Normal Group	<0.001*	<0.001*
	Negative Group	0.022*	0.011*
	Positive Group	0.004*	<0.001*
	Group II	0.902*	0.399*
	Group III	0.899*	0.203*
Group II	Normal Group	<0.001*	<0.001*
	Negative Group	0.029*	0.001*
	Positive Group	0.003*	<0.001*
	Group I	0.902*	0.399*
	Group III	0.998*	0.659*
Group III	Normal Group	<0.001*	<0.001*
	Negative Group	0.029*	<0.001*
	Positive Group	0.003*	0.002*
	Group I	0.899*	0.203*
	Group II	0.988*	0.659*

*Anova Test (Significant <0.05)

Histopathological Picture of the Abdominal Aorta of a Group of Male White Wistar Rats (Rattus



norvegicus) Hyperlipidemia Model After Administration of Kaffir Lime Leaf Ethanol Extract



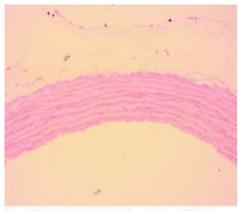
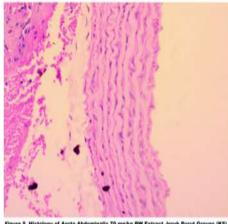


Figure 3. Histology of Aorta Abdominalis Simvastatin Groups (K3)



manification 10x40 (HEE staining).

Discussion

a phytochemical In this study, screening examination of kaffir lime leaf ethanol was carried out and secondary metabolites such as flavonoids, alkaloids, saponins, tannins and phenols were obtained. The results of this examination show that kaffir lime leaves have potential as an antioxidant and anti-inflammatory. The compounds contained in kaffir lime leaves have properties that can be used as traditional medicine [42]. Flavonoid compounds have hydroxy groups that can react with boric acid, producing intense yellow fluorescence at a UV wavelength of 366 nanometers. The results of the alkaloid test showed the formation of an orange precipitate after adding Dragendorff's reagent and a yellow precipitate after adding Mayer's reagent. The presence of saponin can be confirmed by the formation of stable foam after warm water is added. Tannin, on the other hand, is a phenolic compound containing hydroxyl groups, which when reacted with FeCl3 produces a greenish black color.

The results of the analysis of differences in body weight showed that there were significant differences between groups on the 7th and 14th days (p<0.001), but there were no significant differences on the 21st day. These results show that the ethanol extract of kaffir lime leaves can reduce the blood weight of male white Wistar rats with a model of

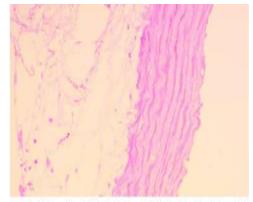
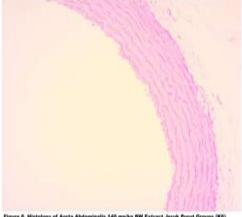


Figure 4. Histology of Aorta Abdominalis 35 mg/kg BW Extract Jeruk Purut Groups (K4) magnification 10x40 (H&E staining).



gure 6. Histology of Aorta Abdominants 140 mg/kg BH Extract Jeruk Furut Groups (Ko) magnification 10x40 (NAE staining).

hyperlipidemia. This can happen because flavonoids can reduce cholesterol levels in the blood by inhibiting the action of the enzyme 3-hydroxy 3methylglutaryl coenzyme A reductase (HMG Co-A reductase) [43]. Reducing cholesterol levels in the bloodstream can reduce fat accumulation in the body's organs and reduce the risk of obesity [44]. By reducing cholesterol levels, the risk of cholesterol buildup in the body's organs decreases, so the risk of obesity is also reduced [45]. Apart from that, compounds such as alkaloids, tannins, saponins, triterpenoids, quinones, steroids, stilbenes, phenolic acids, anthrax, panclin, and hydroxylic acids also play an important role as compounds that can inhibit lipase activity in the body [46].

The results of the analysis of differences in total cholesterol, triglyceride, HDL and LDL levels in male white mice after being given kaffir lime leaf extract showed the effect of disiplidemia. In this study it was seen that there was a decrease in cholesterol levels. This is because kaffir lime leaves contain flavonoid, saponin and tannin compounds which have anti-cholesterol effectiveness (Buathong & Duangsrisai, [47]. Flavonoids can reduce cholesterol levels by inhibiting cholesterol synthesis, increasing LDL receptor expression, and improving cholesterol metabolism overall [48]. As antioxidants, flavonoids can also overcome free

radicals and prevent lipid peroxidation which is an early stage in the formation of atherosclerosis [49]. LDL, as well as increasing the activity of lipoprotein lipase and lecithin cholesterol acyl transferase [50]. The results of the analysis of differences in ENOS levels in this study showed no significant differences between groups in ENOS levels with p>0.05. These results suggest that flavonoids can stimulate eNOS production and increase eNOS expression in the vascular endothelium, which in turn increases NO production. In addition, flavonoids also have antioxidant properties that can protect eNOS and NO from damage by free radicals, maintaining optimal eNOS activity [51]. In the analysis of HMGCOA levels, there was a significant difference between groups in HMGCOA levels in mice (p>0.05). This shows that the compounds contained in the ethanol extract of kaffir lime leaves can inhibit the HMGCOA enzyme [52]. Excessive cholesterol production can be overcome by inhibiting HMG-CoA reductase which mediates the formation of acetyl-CoA into mevalonate; precursor for making cholesterol [53]. Statin drugs have been known to be strong HMG-CoA reductase inhibitors with several undesirable side effects. Therefore, we are looking for safer HMG-CoA reductase inhibitors, especially from kaffir lime leaves [54].

The flavonoids in orange leaf extract are effective in significantly reducing cholesterol and triglyceride levels by inhibiting the enzymes HMG-CoA reductase and acetyl-coenzyme A acetyl-transferase (ACAT) [55]. In their role in reducing blood cholesterol levels in hyperlipidemia conditions, flavonoids can reduce cholesterol synthesis by inhibiting cholesterol esterification in the intestines and liver through inhibiting the activity of the acyl-CoA enzyme cholesterol acyl transferase (ACAT) in HepG2 cells. Apart from that, flavonoids can also inhibit cholesterol synthesis by reducing the activity of the enzyme 3-hydroxyl-3-metal-glutaryl-CoA (HMG-CoA) reductase, thereby reducing cholesterol levels in the liver [56].

Flavonoids function as inhibitors of the HMG-CoA reductase enzyme, thereby causing a decrease in cholesterol synthesis [57]. When cholesterol is transported from the intestine to the liver, the activity of HMG-CoA reductase, which is responsible for converting acetyl-CoA to mevalonate in the process of cholesterol synthesis, is inhibited. As a result, cholesterol production by the liver will be reduced. Alkaloids act as antioxidants by releasing hydrogen ions similar to flavonoids [58]. In addition, this compound can inhibit the activity of the pancreatic lipase enzyme, thereby increasing fat excretion through feces. As a result, fat absorption by the liver is hampered, so fat cannot be converted into cholesterol. Reduced activity of the pancreatic lipase enzyme can reduce the accumulation of triglycerides entering from the small intestine because this enzyme converts triglycerides into two monoglycerides and two free fatty acids which can be absorbed by the blood vessels. In addition, tannins can inhibit fat absorption in the intestine by interacting with mucosal proteins and intestinal epithelial cells [59]. In addition, tannins can accumulate on the protein mucosa on the surface of the small intestine, resulting in a decrease in the effectiveness of cholesterol and fat absorption [60].

4. Conclusions

The results of phytochemical screening on kaffir lime leaves showed secondary metabolites such as flavonoids, phenols, saponins, tannins and alkaloids. In this study there was a significant difference in mean body weight (p < 0.05) on the 14th day but there was no difference between groups on the 21st day. Apart from that, there were no significant differences in mean levels of total cholesterol, triglycerides, HDL and LDL (p<0.05) between all groups. In eNOS levels there were no significant differences (p<0.05) between all groups. However, in HMGCOA levels, there were significant differences in levels (p<0.05) between all groups. Apart from that, the group with the ethanol extract of kaffir lime leaves at a dose of 35 mg/kgBW was better at controlling body weight, reducing total cholesterol, HDL, LDL and triglyceride levels, increasing eNOS levels and reducing HMGCOA levels.

Author Statements:

- **Ethical approval:** The conducted research is not related to either human or animal use.
- **Conflict of interest:** The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper
- Acknowledgement: When conducting research, the author received a lot of support, guidance and direction from various parties, for this reason the author would like to take this opportunity to thank friends, family and lecturers in the Master of Biomedical Sciences Study Program, Faculty of Medicine, Methodist University.
- Author contributions: The authors declare that they have equal right on this paper.
- **Funding information:** The authors declare that there is no funding to be acknowledged.
- **Data availability statement:** The data that support the findings of this study are available on

request from the corresponding author. The data are not publicly available due to privacy or ethical restrictions.

References

- Wang, S., & Bao, X. (2019). Hyperlipidemia, Blood Lipid Levels, and the Risk of Glaucoma: A Meta-Analysis. *Investigative ophthalmology* & visual science, 60(4), 1028-1043. DOI:<u>10.1167/iovs.18-25845</u>
- [2] Abid, R., Mahmood, R., & Santosh Kumar, H.S. (2016). Hypolipidemic and antioxidant effects of ethanol extract of Cassia fistula fruit in hyperlipidemic mice. *Pharmaceutical biology*, 54(12), 2822-2829. <u>https://doi.org/10.1080/13880209.2016.11854</u> <u>45</u>
- [3] Nie, Y., & Luo, F. (2021). Dietary fiber: An for a global control opportunity of hyperlipidemia. Oxidative Medicine and Cellular Longevity, 2021(1),5542342. https://doi.org/10.1155/2021/5542342
- [4] Brechtel, L., Poupore, N., Stoikov, T., Roley, L.T., Emerson, J.F., & Nathaniel, T. (2020). Comorbidities associated with different levels of total cholesterol in male and female acute ischemic stroke patients. *Medicine*, 99(52), e23870. https://doi.org/10.1097/MD.0000000002387

<u>https://doi.org/10.1097/MD.000000000002387</u> 0

[5] Febriani, D., & Besral. (2018). The Effect Of Lifestyle On Hypercholesterolemia. *The Open Public Health Journal*. 2018; 11(1): p. 526– 532.

https://doi.org/10.2174/187494450181101052 6

- [6] Stewart, J., McCallin, T., Martinez, J., Chacko, S., & Yusuf, S. (2020). Hyperlipidemia. *Pediatrics in review*, 41(8), 393-402. <u>https://doi.org/10.1542/pir.2019-0053</u>
- [7] Mortensen, M. B., & Nordestgaard, B. G. (2020). Elevated LDL cholesterol and increased risk myocardial infarction of and atherosclerotic cardiovascular disease in individuals aged 70-100 years: a contemporary primary prevention cohort. The Lancet, 396(10263), 1644-1652. https://doi.org/10.1016/S0140-6736(20):32233-9
- [8] Aguilar-Ballester, M., Herrero-Cervera, A., Vinué, Á., Martínez-Hervás, S., & González-Navarro, H. (2020). Impact of cholesterol metabolism in immune cell function and atherosclerosis. *Nutrients*, 12(7), 2021. <u>https://doi.org/10.3390/nu12072021</u>

- [9] Stone, J. R. (2022). Diseases of small and medium-sized blood vessels. *In Cardiovascular* pathology (pp. 307-351). Academic Press. <u>https://doi.org/10.1016/B978-0-12-822224-9.00020-7</u>
- [10] Lin, C. F., Chang, Y. H., Chien, S. C., Lin, Y. H., & Yeh, H. Y. (2018). Epidemiology of dyslipidemia in the Asia Pacific region. *International Journal of Gerontology*, 12(1), 2-6. <u>https://doi.org/10.1016/j.ijge.2018.02.010</u>
- [11] Summerhill, V. I., Grechko, A. V., Yet, S. F., Sobenin, I. A., & Orekhov, A. N. (2019). The atherogenic role of circulating modified lipids in atherosclerosis. *International journal of molecular sciences*, 20(14), 3561. <u>https://doi.org/10.3390/ijms20143561</u>
- [12] Liu, B. X., Sun, W., & Kong, X. Q. (2019). Perirenal fat: a unique fat pad and potential target for cardiovascular disease. Angiology, 70(7), 584-593. https://doi.org/10.1177/0003319718799967
- [13] Tsao, C. W., Aday, A. W., Almarzooq, Z. I., et al. (2023). Heart disease and stroke statistics 2023 update: a report from the American Heart Association. *Circulation*, 147(8):e93-e621. <u>https://doi.org/10.1161/CIR.00000000000112</u> <u>3</u>
- [14] World Health Organization. Global status report on noncommunicable diseases 2014. <u>http://www.who.int/nmh/publications/ncd-</u> <u>status-report-2014/en/</u>
- [15] Deliara, H., Kartikadewi, A., & Nugraheni, D. M. (2020). Ethanol Extract of Kaffir Lime Peel (Citrus hystrix) Has Potential as a Cholesterol Lowering Agent: In Vivo Study. *Medica Arteriana* (*Med-Art*), 2(1), 1-9. <u>https://doi.org/10.26714/medart.2.1.2020.1-9</u>
- [16] Rokhayati, A., & Rumahorbo, H. (2020). Description of self-efficacy in managing risk factors and maintaining the health function of patients with coronary heart disease. *Bandung Department of Health Polytechnic Health Research Journal*, 12(2), 285-296. <u>https://doi.org/10.34011/juriskesbdg.v12i2.179</u> <u>7</u>
- [17] Hikmah, R. A., Hariadi, P., & Sovia, F. (2022). Effectiveness of infusion of Kepyar Jatropha leaves (Ricinus communis L.) on total cholesterol and HDL blood serum levels of Wistar strain rats. *Sinteza*, 2(1), 86-95. <u>https://doi.org/10.29408/sinteza.v2i1.4367</u>
- [18] Firdaus, AK, Maulana, AR, Laudira, FR, Saputri, NA, Wardan, UZ, & Makky, WR (2022). Education on popular diseases to improve the quality of life for the people of Landungsari Village, Malang Regency. *ABDIMAS: Journal of Community Service*,

Merdeka University, Malang, 7(1), 131-138. https://orcid.org/0000-0003-4761-1565

- [19] He, N., & Ye, H. (2020). Exercise and hyperlipidemia. *Physical exercise for human health*, 79-90. <u>https://doi.org/10.1007/978-981-</u> 15-1792-1 5
- [20] Samuel, PO, Edo, GI, Emakpor, OL, Oloni, GO, Ezekiel, GO, Essaghah, AEA, Agoh, E., & Agbo, JJ (2024). Lifestyle modifications for preventing and managing cardiovascular diseases. *Sport Sciences for Health*, 20(1), 23-36. <u>https://doi.org/10.1007/s11332-023-01118-Z</u>
- [21] Cybulsky, M., Cook, S., Kontsevaya, A.V., Vasiljev, M., & Leon, D.A. (2016).
 Pharmacological treatment of hypertension and hyperlipidemia in Izhevsk, Russia. *BMC Cardiovasc Disord*, 16, 122. <u>https://doi.org/10.1186/s12872-016-0300-9</u>
- [22] Kreisberg, R. A., & Oberman, A. (2003). Medical management of hyperlipidemia/dyslipidemia. *The Journal of Clinical Endocrinology & Metabolism*, 88(6), 2445-2461. <u>https://doi.org/10.1210/jc.2003-030388</u>
- [23] Susanto, K. A., Sitanggang, M. L., Hasan, D., & Chorida, I. (2021). Analysis of Prescription of National Formulary Medicines Based on The Adjustment of Restrictions in Public Hospitals of Sawah Besar 2018. *Indonesian Journal of Pharmaceutical Sciences*, 19(1), 68-72. <u>https://doi.org/10.35814/jifi.v19i1.1056</u>
- [24] Peters, R., Breitner, J., James, S., Jicha, G.A., Meyer, P.F., Richards, M., et al. (2021). Dementia risk reduction: why haven't the pharmacological risk reduction trials worked? An in- depth exploration of seven established risk factors. Alzheimer's å Dementia: **Translational** & Research Clinical Interventions, e12202. 7(1), https://doi.org/10.1002/trc2.12202
- [25] Rauf, A., Akram, M., Anwar, H., et al. (2022) Therapeutic potential of herbal medicine for the management of hyperlipidemia: latest updates. *Environ Sci Pollut Res*, 29, 40281-40301. <u>https://doi.org/10.1007/s11356-022-19733-7</u>
- [26] Gitawati, R., & Widowati, L. (2015). The use of herbal medicine in hyperlipidemia patients is based on medical record data in several health service facilities in Indonesia. *Indonesian Pharmaceutical Journal*, 5(1), 41-48. <u>https://doi.org/10.22435/jki.v5i1.3474</u>
- [27] Hasani-Ranjbar, S., Nayebi, N., Moradi, L., Mehri, A., Larijani, B., & Abdollahi, M. (2010). The efficacy and safety of herbal medicines used in the treatment of hyperlipidemia; a systematic review. *Current pharmaceutical*

design, 16(26), 2935-2947. <u>https://doi.org/10.2174/138161210793176464</u>

- [28] Tirawanchai, N., Kengkoom, K., Isarangkul, D., Burana-Osot, J., Kanjanapruthipong, T., Chantip, S., et al. (2020). A combination extract of kaffir lime, galangal, and lemongrass maintains blood lipid profiles, hepatocytes, and liver mitochondria in rats with nonalcoholic steatohepatitis. *Biomedicine & Pharmacotherapy*, 124, 109843. <u>https://doi.org/10.1016/j.biopha.2020.109843</u>
- [29] Erba, O., Atomsa, D., Chimdessa, M., Gonfa, T. (2020). Determination of Flavonoid Contents and Evaluation of in vitro Antioxidant Activities of the Extract of Selected Citrus Fruit Peel. *International Journal of Secondary Metabolites*, 7(1), 8-18. https://doi.org/10.21448/ijsm.660578
- [30] Kumar, R., Mishra, S., Shekher, R., & Sharma, R. K. (2023, November). Exploration and Characterization of the Diverse Bioactive Constituents Present in Citrus Fruit Peels: An In-Depth Study on Their Functional Properties and Potential Health Benefits. *In 2023 International Conference on Integration of Computational Intelligent Systems (ICICIS)*. (pp. 1-5). IEEE. DOI: 10.1109/ICICIS56802.2023.10430268
- [31] Hu, Y., Xu, J., Chen, Q., Liu, M., Wang, S., Yu, H., et al. (2020). Regulatory effects of total flavonoids in Morus alba L. on hepatic cholesterol disorders in orotic acid induced NAFLD rats. *BMC Complementary Medicine* and Therapies, 20:257, 1-12. https://doi.org/10.1186/s12906-020-03052-w
- [32] Siti, H. N., Mohamed, S., & Kamisah, Y. (2022). Potential therapeutic effects of Citrus hystrix DC and its bioactive compounds on metabolic disorders. *Pharmaceuticals*, 15(2), <u>167. https://doi.org/10.3390/ph15020167</u>
- [33] Ullah, A., Munir, S., Badshah, S. L., Khan, N., Ghani, L., et al. (2020). Important flavonoids and their role as a therapeutic agent. *Molecules*, 25(22), 5243. https://doi.org/10.3390/molecules25225243
- [34] Ito, F. (2021). Polyphenols can potentially prevent atherosclerosis and cardiovascular disease by modulating macrophage cholesterol metabolism. *Current Molecular Pharmacology*, 14(2), 175-190. <u>https://doi.org/10.2174/187446721366620032</u> 0153410
- [35] Soltani, S., Boozari, M., Cicero, A.F., Jamialahmadi, T., & Sahebkar, A. (2021).
 Effects of phytochemicals on macrophage cholesterol efflux capacity: Impact on

atherosclerosis. *Phytotherapy Research*, 35(6), 2854-2878. https://doi.org/10.1002/ptr.6991

- [36] Rustanti, E., & Lathifah, Q. A. Y. (2018). Identifikasi senyawa kuersetin dari fraksi etil asetat ekstrak daun alpukat (persea americana mill.). *Alchemy: Journal of Chemistry*, 6(2), 38-42. <u>https://doi.org/10.18860/al.v6i2.6768</u>
- [37] Shafreen, R. B., Lubinska-Szczygeł, M., Różańska, A., Dymerski, T., Namieśnik, J., Katrich, E., & Gorinstein, S. (2018). Human serum interactions with phenolic and aroma substances of Kaffir (Citrus hystrix) and Key lime (Citrus aurantifolia) juices. *Journal of Luminescence*, 201, 115-122. https://doi.org/10.1016/j.jlumin.2018.04.010
- [38] Shutler, S. M., Walker, A. F., & Low, A. G. (1987). The Cholesterol-Lowering Effects of Legumes Ii: Effects of Fiber, Sterols, Saponins and Isolavones. Human Nutrition. *Food Sciences and Nutrition*, 41(2), 87-102. https://doi.org/10.1080/09528954.1987.11904 104
- [39] Afrose, S., Hossain, M. S., Maki, T., & Tsujii, H. (2009). Karaya root saponin exerts a hypocholesterolemic response in rats fed a high-cholesterol diet. *Nutrition research*, 29(5), 350-354.

https://doi.org/10.1016/j.nutres.2009.05.008

[40] Zeng, X., Jiang, W., Du, Z., & Kokini, J. L. (2023). Encapsulation of tannins and tanninrich plant extracts by complex coacervation to improve their physicochemical properties and biological activities: A review. *Critical Reviews in Food Science and Nutrition*, 63(18), 3005-3018.

https://doi.org/10.1080/10408398.2022.20753 13

[41] Djabir, Y. Y., Pratiwi, Y., & Bahar, M. A. (2022). Preclinical Study of the Effects of Simvastatin, Rosuvastatin, and Fenofibrate on Lactate Dehydrogenase Levels in Female Rats Induced by Oral Contraception and a High-Fat Diet. *Pharmacy and Pharmacology Magazine*, 26(1), 4-7. https://doi.org/10.2005/.mff.v26i1.18125

https://doi.org/10.20956/mff.v26i1.18125

- [42] Suresh, A., Velusamy, S., Ayyasamy, S., & Rathinasamy, M. (2021). Techniques for essential oil extraction from kaffir lime and its application in health care products—A review. Flavor and Fragrance Journal, 36(1), 5-21. <u>https://doi.org/10.1002/ffj.3626</u>
- [43] Ranti, G. C., Fatimawali, F., & Wehantouw, F. (2013). Test the effectiveness of flavonoid and steroid extracts from Gedi (abelmoschus manihot) as anti-obesity and hypolipidemic in male white Wistar rats. *Pharmacon*, 2(2), 34-38. <u>https://doi.org/10.35799/pha.2.2013.1554.</u>

[44] Song, Y., Liu, J., Zhao, K., Gao, L., & Zhao, J. (2021). Cholesterol-induced toxicity: An integrated view of the role of cholesterol in multiple diseases. *Cell metabolism*, 33(10), 1911-1925.

https://doi.org/10.1016/j.cmet.2021.09.001

- [45] Després, J. P., Arsenault, B. J., Côté, M., Cartier, A., & Lemieux, I. (2008). Abdominal obesity: the cholesterol of the 21st century?. *Canadian Journal of Cardiology*, 24, 7D-12D. <u>https://doi.org/10.1016/S0828-</u> 282X(08)71043-2
- [46] Ujiana, W. O., Meak, L. E. C., Hiko, M. F., & Bare, Y. (2022). Potential of Triterpenoid Compounds in Sauropus androgynus L Merr as In Silico Inhibitor of Obesity. *Spizaetus: Jurnal Biologi dan Pendidikan Biologi*, 3(2), 43-47. <u>https://doi.org/10.55241/spibio.v3i2.64</u>
- [47] Buathong, R., & Duangsrisai, S. (2023). Plant ingredients in Thai food: a well-rounded diet for natural bioactives associated with medicinal properties. *PeerJ*, 11, e14568. <u>https://doi.org/10.7717/peerj.14568</u>
- [48] Millar, C. L., Duclos, Q., & Blesso, C. N. (2017). Effects of dietary flavonoids on reverse cholesterol transport, HDL metabolism, and HDL function. *Advances in Nutrition*, 8(2), 226-239.

https://doi.org/10.3945/an.116.014050

- [49] Ilyasov, I., Beloborodov, V., Antonov, D., Dubrovskaya, A., Terekhov, R., Zhevlakova., et al. (2020). Flavonoids with glutathione antioxidant synergy: Influence of free radicals inflow. *Antioxidants*, 9(8), 695. https://doi.org/10.3390/antiox9080695
- [50] Saputra, I. P. B. A., & Arjita, I. P. D. (2024). The Potential of Flavonoid Derivative Compounds as Inhibitors of the HMG-CoA Reductase Enzyme for Candidates of Hypercholesterolemia Drugs. *Journal of Science Education Research*, 10(5), 2286-2293. <u>https://doi.org/10.29303/jppipa.v10i5.6237</u>
- [51] Duarte, J., Francisco, V., & Perez-Vizcaino, F. (2014). Modulation of nitric oxide by flavonoids. *Food & function*, 5(8), 1653-1668. <u>https://doi.org/10.1039/C4FO00144C</u>
- [52] Ademosun, A. O., Oboh, G., Olasehinde, T. A., & Adeoyo, O. O. (2018). From folk medicine to functional food: A review on the bioactive components and pharmacological properties of citrus peels. Oriental Pharmacy and Experimental Medicine, 18, 9-20. https://doi.org/10.1007/s13596-017-0292-8
- [53] Su, X. Z., Zhang, L. F., Hu, K., An, Y., Zhang, Q. P., Tang, J. W., et al. (2024). Discovery of Natural Potent HMG- CoA Reductase Degraders for Lowering Cholesterol.

Angewandte Chemie International Edition, 63(6), e202313859. https://doi.org/10.1002/anie.202313859

- [54] Santosa, R. A., Yuna, U. N., & Virgana, R. (2021). Effectiveness of Statins in Preventing the Onset and Progression of Age Related Macular Degeneration: A Systematic Review. Journal of Ophthalmology, 3(1), 35-47. https://doi.org/10.11594/ojkmi.v3i1.12
- [55] Depari, S. A. F., Rambe, D. J. A., Meilando, R., Lisya, C., Mutia, M. S., & Lubis, Y. E. P. (2021). -Effectiveness Test of Sunkist Orange Peel Ethanol Extract (Citrus sinensis (L.) Osbeck) on Blood Sugar Levels of Wistar Rats norvegicus) with Streptozotocin-(Rattus Induced Hypercholesterolemia. Biospecies, 14(1),1-9. https://doi.org/10.22437/biospecies.v14i1.1125
- [56] Iqbal, M., Ricky, N., Indra, T., & Muhammad, B. (2021). Potential of Karamunting Flower (Melastoma Malabathricum L.) on Total Cholesterol and Triglyceride Levels in Male White Rats with Propylthiouracil-Induced Hyperlipidemia. Achievement Journal. 14(1):

https://doi.org/10.22437/biospecies.v14i1.1125 0.

[57] Baskaran, G., Salvamani, S., Ahmad, S. A., Shaharuddin, N. A., Pattiram, P. D., & Shukor, M. Y. (2015). HMG-CoA reductase inhibitory activity and phytocomponent investigation of Basella alba leaf extract as a treatment for hypercholesterolemia. Drug design, 509-517. development and therapy, https://doi.org/10.2147/DDDT.S75056

[58] Pratiwi, L., Fudholi, A., Martien, R., & Pramono, S. (2016). Ethanol extract, ethyl acetate extract, ethyl acetate fraction, and nhexane fraction mangosteen peels (Garcinia mangostana L.) as source of bioactive substance free-radical scavengers. JPSCR: Journal of Pharmaceutical Science and Clinical Research, 1(2), 71-82. https://doi.org/10.20961/jpscr.v1i2.1936

[59] Kardum, N., & Glibetic, M. (2018). Polyphenols and their interactions with other dietary compounds: Implications for human health. In Advances in food and nutrition research, (Vol. 84, pp. 103-144). Academic Press.

https://doi.org/10.1016/bs.afnr.2017.12.001

[60] Artha, C., Mustika, A., & Sulistyawati, S. W. (2017). Effect of singawalang leaf extract on LDL levels of hypercholesterolemic male white mice. Indonesian Medical eJournal, 5(2), 105-109. https://doi.org/10.23886/ejki.5.7151